

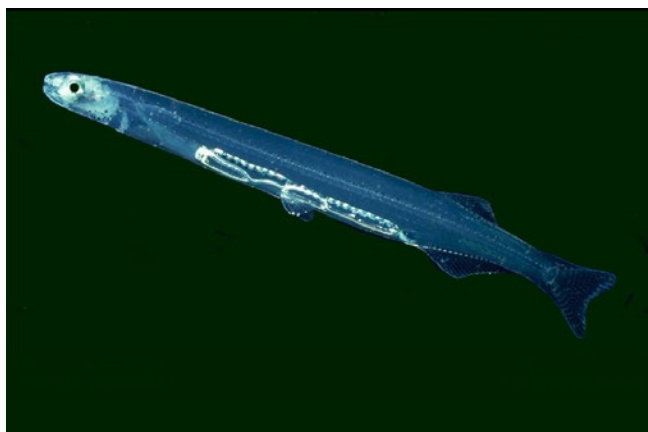
IMPACTS OF SEDIMENT ON ĪNANGA

Sediment can affect māhinga kai by influencing habitat, behaviour, feeding, growth and survival.

Background on Īnanga (*Galaxias maculatus*)

Īnanga are one of six species in New Zealand's whitebait catch. They make up over 87% of the whitebait caught in rivers around the whole country¹. Īnanga are diadromous - they spend half their life in the ocean as larvae and the other half in rivers as juveniles and adults². They mature after re-entering freshwater, but unlike salmon, most do not return to the river where they were born³. Maturing Īnanga have very general habitat requirements⁴. They are more active during daylight, forming large shoals in a wide range of coastal waterways⁴. Īnanga feed on a broad range of aquatic⁵⁻⁷ and terrestrially-derived⁸ food. Most adults only live for one year and spawn once before dying⁹. Females lay up to 4,000 eggs⁹ in riverbank vegetation while it is submerged during spring (full and new moon) high tides. The eggs need dense vegetation to protect them from temperature extremes and from drying out while they develop¹⁰. Īnanga also occur in Australia (where they are called 'common jollytail') and South America (where they are called 'puye').

Īnanga juvenile (*Galaxias maculatus*)



Īnanga sensitivity to elevated sediment



Low Medium High

IMPACTS OF SEDIMENT ON ĪNANGA

Effects of suspended sediment on Īnanga

Habitat Direct effects unknown.

Behaviour Īnanga are mobile and do not have territories, so their likely response to a change in water quality will be avoidance if possible. Īnanga whitebait will not swim into very turbid water¹¹. However, it is unlikely that this level of turbidity would occur for a long enough period during whitebait migrations to reduce the overall number of juvenile Īnanga that enter a river. Adult Īnanga that are already living in a river may move to areas of less turbid water (e.g., backwaters or among bank vegetation) during a flood event if they are available. This behaviour may allow them to avoid turbid water, but it may also increase their risk of predation or reduce their ability to access food.

Feeding Īnanga are highly dependent on sight for feeding, but their ability to see food is not reduced significantly until very high turbidity levels¹². They are probably more visually sensitive than other whitebait species because of their large eyes¹³ and optic lobes¹⁴. Feeding in juvenile Īnanga reduces when turbidity reaches very high levels¹², this reduced feeding rate comes directly through a reduction in their ability to feed rather than indirectly through a stress-related reduction in appetite¹². Adult Īnanga show no change in feeding rate at high suspended sediment levels¹⁵.

Growth Sustained periods (21 days) of moderate levels of turbidity reduce the growth of juvenile Īnanga¹⁶. This is probably an indirect effect caused by reduced feeding efficiency.

Survival Repeated, short-term (< 24 hours) exposure to very high turbidity does not reduce the survival of juvenile Īnanga¹⁷. Even long-term (21 days) exposure to very high turbidity has no effect on survival¹⁶. Studies have shown that turbidity levels have to reach around 20,000 NTU to cause 50% of juvenile Īnanga to die and 30,000 NTU to cause 100% mortality¹⁷. The cause of death is likely sediment damaging their gills. In the wild, other conditions may occur at the same time as high turbidity, like increased water acidity or low oxygen levels. Together these conditions might mean reduced fish survival rates.

Effects of deposited sediment on Īnanga

Habitat Īnanga live their lives in open water, so sediments deposited on the bottom of rivers are unlikely to impact them directly. However, for spawning, Īnanga use habitats that are very vulnerable to deposited sediments¹⁸. Īnanga eggs need the protective microclimate underneath riparian vegetation to survive¹⁰. If deposited sediments clog the aerial root mats under the riparian vegetation, the humidity around the developing eggs will decrease and the eggs will die¹⁹. Sediment deposited in Īnanga spawning sites during flood events can bury and kill developing eggs. The surface layer of Īnanga eggs is sticky to help them adhere to riparian vegetation²⁰; this prevents the eggs being washed downstream. However, the adhesive layer does cause the eggs to become coated with sediment and this may reduce oxygen transfer to the embryo developing inside²¹.

Behaviour Direct effects unknown.

Feeding Īnanga are mid-depth feeders and avoid feeding on the substrate⁷, so deposited sediments probably won't directly affect their feeding. However, deposited sediment will probably reduce the abundance of their common food sources (e.g., chironomids)²²; Īnanga feed on these insects as they drift downstream with the currents to colonise new habitats.

Growth Any indirect reduction in feeding, through deposited sediment reducing the abundance of food sources, is likely to impact the growth of Īnanga. However, they are opportunistic and mobile feeders so it is possible that they can switch food sources or move to unaffected areas if they are available. Īnanga rely heavily on drift feeding, so it is possible that adequate food supplies may still drift into areas affected by localised sediment deposition from unaffected areas upstream.

Survival The survival of Īnanga will decrease if the amount of food available is reduced for extended periods by deposited sediment.

IMPACTS OF SEDIMENT ON ĪNANGA

Further information:

1. Yungnickel, M.R., M.J.H. Hickford, and D.R. Schiel (2020). Spatio-temporal variation in species composition of New Zealand's whitebait fishery. *New Zealand Journal of Marine and Freshwater Research* 54(4): 679-694.
2. Egan, E.M.C., M.J.H. Hickford, and D.R. Schiel (2019). Understanding the life histories of amphidromous fish by integrating otolith-derived growth reconstructions, post-larval migrations and reproductive traits. *Aquatic Conservation: Marine and Freshwater Ecosystems* 29(9): 1391-1402.
3. Hickford, M.J.H. and D.R. Schiel (2016). Otolith microchemistry of the amphidromous *Galaxias maculatus* shows recruitment to coastal rivers from unstructured larval pools. *Marine Ecology Progress Series* 548: 197-207.
4. Sagar, P.M. (1993). Habitat use and models of abundance of maturing inanga in South Island, New Zealand, streams. *New Zealand Freshwater Miscellaneous Report* 104: 1-29.
5. Catlin, A.K., K.J. Collier, and I.C. Duggan (2019). Diet of juvenile *Galaxias maculatus* (Galaxiidae) during the upstream migration period in the lower Waikato River, New Zealand. *Marine and Freshwater Research* 70(6): 816-823.
6. Stuart, R.E., T. Ingram, and G.P. Closs (2021). Growth and diet of inanga (*Galaxias maculatus*) within a small New Zealand coastal pond system. *New Zealand Journal of Marine and Freshwater Research*: 1-16.
7. Richardson, J., D.W. West, and G. Croker (1997). Who ordered the Austrosimulium on Egeria? *Water & Atmosphere* 5(3): 14-16.
8. Jowett, I.G. (2002). In-stream habitat suitability criteria for feeding inanga (*Galaxias maculatus*). *New Zealand Journal of Marine and Freshwater Research* 36(2): 399-407.
9. Stevens, J.C.B., M.J.H. Hickford, and D.R. Schiel (2016). Evidence of iteroparity in the widely distributed diadromous fish inanga *Galaxias maculatus* and potential implications for reproductive output. *Journal of Fish Biology* 89(4): 1931-1946.
10. Hickford, M.J.H. and D.R. Schiel (2011). Synergistic interactions within disturbed habitats between temperature, relative humidity and UVB radiation on egg survival in a diadromous fish. *PLoS ONE* 6(9): e24318.
11. Boubée, J.A.T., T.L. Dean, D.W. West, and R.F.G. Barrier (1997). Avoidance of suspended sediment by the juvenile migratory stage of six New Zealand native fish species. *New Zealand Journal of Marine and Freshwater Research* 31(1): 61-69.
12. Rowe, D.K. and T.L. Dean (1998). Effects of turbidity on the feeding ability of the juvenile migrant stage of six New Zealand freshwater fish species. *New Zealand Journal of Marine and Freshwater Research* 32(1): 21-29.
13. McDowall, R.M. (1997). An accessory lateral line in some New Zealand and Australian galaxiids (Teleostei: Galaxiidae). *Ecology of Freshwater Fish* 6(4): 217-224.
14. Cadwallader, P.L. (1975). Relationship between brain morphology and ecology in New Zealand Galaxiidae, particularly *Galaxias vulgaris* (Pisces: Salmoniformes). *New Zealand Journal of Zoology* 2(1): 35-43.
15. Rowe, D.K., J.P. Smith, and E. Williams (2002). Effects of turbidity on the feeding ability of adult, riverine smelt (*Retropinna retropinna*) and inanga (*Galaxias maculatus*). *New Zealand Journal of Marine and Freshwater Research* 36(1): 143-150.
16. Cavanagh, J.E., K.L. Hogsden, and J.S. Harding (2014). Effects of suspended sediment on freshwater fish, in Landcare Research Contract Report No. LC1986, 2p.
17. Rowe, D.K., et al. (2002). Lethal turbidity levels for common freshwater fish and invertebrates in Auckland streams, in NIWA Client Report ARC02283. Prepared for Auckland Regional Council: Auckland 37p.
18. Hickford, M.J.H. and D.R. Schiel (2011). Population sinks resulting from degraded habitats of an obligate life-history pathway. *Oecologia* 166(1): 131-140.
19. Hickford, M.J.H. and D.R. Schiel (2014). Experimental rehabilitation of degraded spawning habitat of a diadromous fish, *Galaxias maculatus* (Jenyns, 1842) in rural and urban streams. *Restoration Ecology* 22(3): 319-326.
20. Benzie, V. (1968). Stages in the normal development of *Galaxias maculatus attenuatus* (Jenyns). *New Zealand Journal of Marine and Freshwater Research* 2(4): 606-627.
21. Benzie, V. (1968). Some ecological aspects of the spawning behaviour and early development of the common whitebait *Galaxias maculatus attenuatus* (Jenyns). *Proceedings of the New Zealand Ecological Society* 15: 31-39.
22. Franklin, P.A., et al. (2019). Deriving potential fine sediment attribute thresholds for the National Objectives Framework, in NIWA Client Report 2019039HN. Prepared for Ministry for the Environment: Hamilton 290p.

Prepared by Mike Hickford, Michele Melchior and Melanie Mayall-Nahi from NIWA for Our Land and Water National Science Challenge, March 2023. Image of inanga whitebait by Dr R M McDowall.